

GPAT Pharmaceutics

Sample Paper – 9

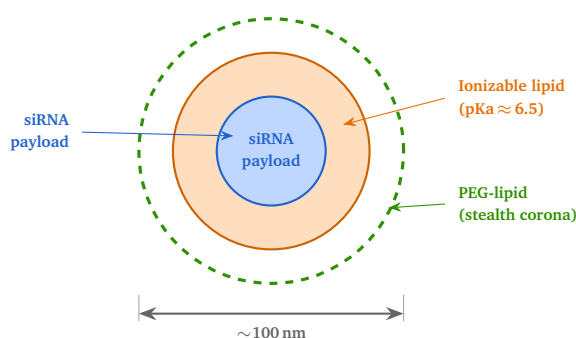
Duration: 36 Minutes

Maximum Marks: 100

Instructions

- This paper contains **25** Multiple Choice Questions (Single Correct Answer), modelled on the Pharmaceutics portion of **GPAT** entrance.
- Each correct answer carries **+4 marks**. There is a **penalty of –1 mark** for each incorrect answer. Unattempted questions carry no marks.
- Only **one** option is correct per question.
- Syllabus: **GPAT Pharmaceutics — Novel Drug Delivery Systems (NDDS), Biotechnology Pharmaceuticals, Regulatory Affairs.**
- Personal calculators, mobile phones, and electronic gadgets are strictly prohibited. A simple on-screen calculator is provided in the CBT interface; rough sheets are supplied at the centre.

Q1. The figure below shows a cross-sectional diagram of an siRNA lipid nanoparticle (LNP) containing DLin-MC3-DMA (MC3) as the ionizable lipid ($pK_a \approx 6.5$).



The **PRIMARY** reason for selecting an ionizable lipid with $pK_a \approx 6.5$ for systemic intravenous siRNA delivery is:

- (A) At physiological pH 7.4 the lipid is nearly neutral (uncharged), minimising serum protein binding and RBC interaction; at endosomal



pH $\approx$ 5.5 (post-endocytosis) it becomes cationic, disrupting the endosomal membrane to enable siRNA cytoplasmic escape

- (B) The lipid remains permanently cationic at all physiological pH values, ensuring continuous electrostatic binding to the anionic siRNA cargo throughout the circulation and after endocytosis
- (C) A pK_a of 6.5 ensures the ionizable lipid is maximally protonated at pH 7.4, optimising LNP cellular uptake by electrostatic interaction with the negatively charged plasma membrane
- (D) The lipid undergoes irreversible acid-catalysed hydrolysis at endosomal pH 5.5, releasing the siRNA payload by chemical degradation of ester bonds in the LNP core

Q2. Abraxane (nab-paclitaxel) is formulated as albumin-bound paclitaxel nanoparticles ($\sim$ 130 nm). Compared to conventional Cremophor EL-solubilised paclitaxel (Taxol), what is the **PRINCIPAL** mechanism by which Abraxane achieves improved tumour accumulation?

- (A) Enhanced passive EPR (enhanced permeability and retention) effect arising solely from the smaller 130 nm particle size compared to Cremophor EL micelles (<10 nm)
- (B) Albumin-mediated active transcytosis across tumour endothelial cells via the gp60 receptor (caveolin-1-dependent), followed by capture of albumin-drug by SPARC (secreted protein acidic and rich in cysteine) overexpressed in the tumour stroma
- (C) Elimination of Cremophor EL prevents P-glycoprotein-mediated efflux, allowing greater intracellular drug retention in tumour cells without any active tumour targeting mechanism
- (D) The albumin coat activates Fc γ receptor-expressing tumour-infiltrating macrophages, causing preferential phagocytosis of Abraxane inside the tumour microenvironment

Q3. In antibody-drug conjugate (ADC) development, the drug-to-antibody ratio (DAR) is a critical quality attribute. What is the **MAJOR pharmacokinetic consequence** of a DAR greater than 8?



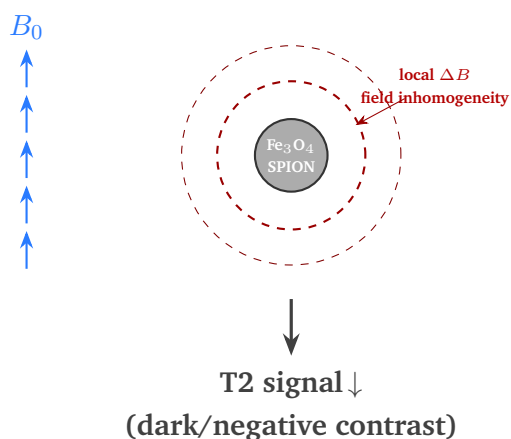
- (A) Reduced cytotoxicity, because excess drug molecules sterically block antigen binding and prevent the ADC from reaching the target receptor on tumour cells
- (B) Greatly enhanced antigen-binding affinity, leading to a hook effect that paradoxically saturates tumour receptors and prevents internalisation
- (C) Rapid systemic clearance due to hydrophobic aggregation of the over-loaded ADC, combined with increased off-target toxicity from premature payload release in non-tumour tissues
- (D) Permanent loss of antibody-dependent cellular cytotoxicity (ADCC) because the Fc domain is sterically blocked by drug-linker units conjugated to lysine residues adjacent to the Fc region

Q4. CdSe/ZnS core-shell quantum dots (QDs) are under investigation as fluorescent imaging probes. The **PRIMARY** source of cytotoxicity associated with CdSe/ZnS quantum dots is:

- (A) The ZnS shell reacts with intracellular glutathione to form toxic zinc-thiolate complexes that inhibit mitochondrial complex I in the electron transport chain
- (B) The large surface-area-to-volume ratio causes non-specific adsorption of all plasma proteins simultaneously, triggering complement-mediated anaphylaxis
- (C) The quantum confinement effect generates gamma (γ) radiation inside cells, causing direct DNA double-strand breaks and irreversible genomic instability
- (D) Release of free Cd^{2+} ions from the CdSe core via photooxidation, mechanical stress, or intracellular metabolic degradation, generating reactive oxygen species and inhibiting mitochondrial function

Q5. The figure below illustrates a superparamagnetic iron oxide nanoparticle (SPION) placed in an external magnetic field B_0 .





SPIONs used as MRI contrast agents produce T2-weighted **negative** contrast by:

- (A) Generating heat via Néel and Brownian relaxation that directly destroys adjacent water molecules, eliminating their contribution to the MRI signal
- (B) Creating local magnetic field inhomogeneities around each nanoparticle that accelerate T2/T2* relaxation of surrounding water protons, producing signal loss (dark contrast) on T2-weighted images
- (C) Directly absorbing the radiofrequency excitation pulses, preventing surrounding tissue protons from being tipped into the transverse plane and producing signal
- (D) Permanently aligning water proton spins parallel to B_0 in a ferromagnetic state, eliminating transverse magnetisation and generating uniform signal voids throughout the entire tissue volume

Q6. Poly(*N*-isopropylacrylamide) (PNIPAM) is formulated as a thermoresponsive injectable hydrogel with an LCST of 34 °C. The formulation is administered subcutaneously at room temperature (25 °C) and is intended to gel in situ at body temperature (37 °C). Which description **CORRECTLY** characterises the behaviour of PNIPAM **BELOW** its LCST?

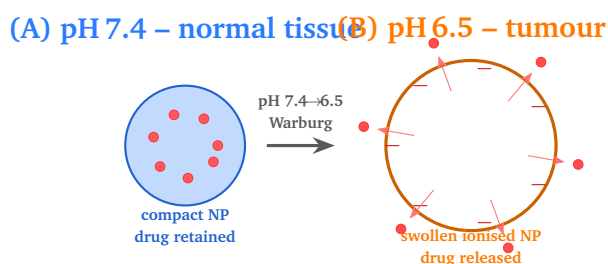
- (A) The polymer chain is hydrophilic and adopts an extended, water-swollen coil conformation; the solution is free-flowing and injectable at room temperature below 34 °C
- (B) The polymer collapses into a compact hydrophobic globule, expelling



water and forming a rigid non-injectable gel at room temperature below 34 °C

- (C) PNIPAM undergoes irreversible covalent cross-linking via amine–carboxylate condensation below 34 °C, permanently gelling the solution before administration
- (D) PNIPAM is fully insoluble in water below its LCST and must be dissolved in an organic co-solvent (e.g., ethanol) before subcutaneous injection

Q7. The diagram below shows the behaviour of a pH-responsive polymer nanoparticle (poly-methacrylic acid, PMAA) in normal tissue and in the tumour microenvironment.



pH-responsive polymers exploit the Warburg effect in tumour targeting by:

- (A) Detecting elevated alkaline phosphatase in the tumour, which raises intratumoral pH to ~8.0; the polymer dissolves only at this alkaline pH, releasing drug exclusively in tumours
- (B) Being protonated and uncharged at blood pH 7.4 and undergoing spontaneous hydrolysis at any pH < 7.4, including the stomach, causing non-selective premature drug release in the gastrointestinal tract
- (C) Taking advantage of excess lactic acid generated by anaerobic glycolysis (Warburg effect) in tumour cells, which reduces the extracellular tumour pH to 6.4–6.8; ionisation of the polymer's acidic groups at this pH causes chain expansion and drug release into the tumour interstitium
- (D) Responding to elevated intracellular glutathione in the tumour cytoplasm by cleaving disulfide cross-links, releasing drug intracellularly rather than in the extracellular tumour microenvironment



Q8. A pulsatile tablet for nocturnal asthma uses an erodible outer coating (hydroxylated ethylcellulose) to generate a 6-hour lag time before releasing a burst dose of salbutamol. The tablet is taken at bedtime (~ 10 pm). What is the **PRIMARY pharmacological rationale** for programming this specific 6-hour lag time?

- (A) The lag time prevents contact between salbutamol and gastric acid; drug is released only once the erodible coat has carried the tablet into the alkaline duodenum for absorption
- (B) A 6-hour delay allows the tablet to transit to the colon where colonic azoreductase cleaves the coating, enabling targeted colonic absorption of salbutamol to act on the enteric nervous system
- (C) Salbutamol has a half-life of ~ 30 minutes; the lag time allows the initial dose to be fully eliminated before the pulsatile dose is released, preventing accumulation and arrhythmia
- (D) Peak bronchoconstriction and the nadir of pulmonary function in nocturnal asthma occur in the early morning (3–5 am); a 6-hour lag after bedtime administration ($10 \text{ pm} + 6 \text{ h} = 4 \text{ am}$) synchronises peak salbutamol plasma levels with this circadian symptomatic peak

Q9. Gastroretentive pellets are formulated by incorporating barium sulphate (BaSO_4 , density = 4.48 g/cm^3) to achieve a pellet density of 2.7 g/cm^3 . Gastric fluid has a density of approximately 1.004 g/cm^3 . The **MECHANISM** by which these high-density pellets are retained in the stomach is:

- (A) The pellets are paramagnetic and adhere to the antral wall via an externally applied magnetic field; BaSO_4 provides the superparamagnetic properties required for gastric mucoadhesion
- (B) Being denser than gastric contents, the pellets sediment to the most dependent region (antrum/lower curvature) of the stomach, where they are retained by gravity and the anatomy of the pyloric canal, avoiding the peristaltic wave that empties lighter floating objects
- (C) BaSO_4 reacts with gastric HCl to release Ba^{2+} ions, which cross-link gastric mucin and form an adhesive gel that immobilises the pellets



at the gastric mucosa

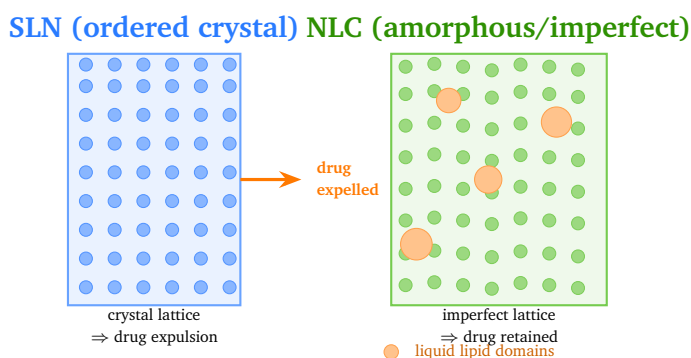
- (D) The high density generates an elevated osmotic pressure inside the pellet matrix that drives gastric fluid into the core, causing the pellet to swell beyond the pyloric sphincter diameter and block gastric emptying

Q10. Thiomers (thiolated polymers, e.g., chitosan-cysteine, polycarbophil-cysteine) exhibit superior mucoadhesion compared to conventional non-thiolated polymers such as Carbopol and hydroxyethyl cellulose (HEC). The reason thiomers outperform conventional mucoadhesive polymers is that they:

- (A) Form covalent disulfide bonds ($-S-S-$) with free cysteine residues in the Cys-rich subdomains of mucin glycoproteins, providing chemical (covalent) mucoadhesion in addition to physical chain entanglement
- (B) Chelate calcium ions from the mucosal surface to form stable calcium-thiolate bridges that mechanically anchor the dosage form to the mucosal epithelium
- (C) React with histidine residues in mucin via acid-base reactions at physiological pH, forming salt bridges that are 100-fold stronger than conventional hydrogen bonds
- (D) Expand tight junctions between mucosal epithelial cells by denaturing claudin and occludin proteins through thiol-disulfide exchange reactions

Q11. The figure below compares the internal architecture of a solid lipid nanoparticle (SLN) and a nanostructured lipid carrier (NLC).





Nanostructured lipid carriers (NLC) overcome the main limitation of SLNs by:

- (A) Replacing the lipid matrix entirely with a biodegradable PLGA polymer that does not undergo crystalline transitions and therefore retains drug indefinitely without expulsion
- (B) Adding a second ionic surfactant layer around the SLN core that acts as an additional drug reservoir, increasing total payload by electrostatic interaction
- (C) Blending a solid lipid with a spatially incompatible liquid lipid to create an imperfect, amorphous crystal structure; this prevents the transition to an ordered β -crystal lattice that otherwise expels drug molecules during SLN storage
- (D) Using exclusively solvent emulsification-evaporation preparation, which produces particles < 50 nm with inherently better physical stability than hot-homogenisation (HPH)-prepared SLNs

Q12. Transfersomes (elastic/deformable vesicles) contain an edge activator (e.g., sodium cholate, Tween 80) that makes the bilayer sufficiently flexible to squeeze through skin intercellular channels much narrower than the vesicle diameter (~ 200 – 300 nm). They must be applied **non-occlusively** (open skin surface, not under an impermeable patch). The **PRIMARY** reason for the non-occlusive application requirement is:

- (A) Occlusion traps residual organic solvent (ethanol) from the vesicle preparation within the stratum corneum, causing chemical burns that open-application prevents by allowing solvent evaporation



- (B) Occlusion prevents the edge activator from desorbing from the bilayer; without periodic loss of sodium cholate into the skin surface, the bilayer remains too rigid to deform and penetrate
- (C) Under occlusion, transferosome lipids fuse with sebaceous gland secretions, forming stable lipid plugs that permanently block sebaceous gland ducts and halt vesicle migration
- (D) The transepidermal water gradient (higher water activity inside the viable epidermis than at the skin surface) drives vesicle migration inward; occlusion eliminates surface evaporation and thereby abolishes this gradient, removing the thermodynamic driving force for transferosome penetration

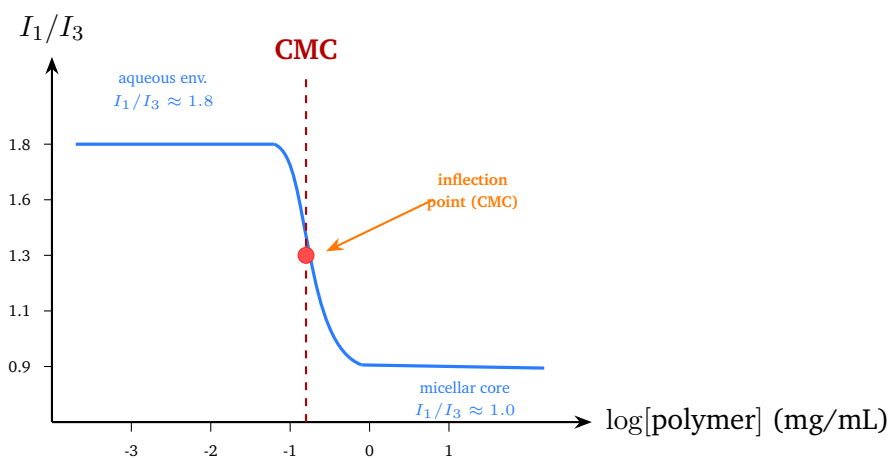
Q13. Exosomes (30–150 nm endogenous extracellular vesicles derived from multivesicular bodies) are being investigated as alternatives to synthetic liposomes for drug delivery. Compared to synthetic liposomes, what is the **PRIMARY BIOLOGICAL ADVANTAGE** of exosomes as drug delivery vehicles?

- (A) Exosomes are far easier to manufacture at industrial scale, with uniform size distribution and substantially higher drug loading efficiency than conventional thin-film hydration liposomes
- (B) Because exosomes are derived from cell membranes, they display endogenous surface proteins (integrins, tetraspanins, CD47 “don’t-eat-me” signal) that confer natural cellular homing ability and evade immune recognition, resulting in intrinsically lower immunogenicity than synthetic vesicles
- (C) Exosome membranes are composed entirely of synthetic PEGylated lipids, providing superior steric stabilisation and longer circulation half-life than conventional liposomes that lack PEG
- (D) The internal aqueous lumen of exosomes is maintained at pH 5.0, enabling encapsulation of pH-sensitive drugs without degradation under physiological conditions

Q14. The critical micelle concentration (CMC) of an amphiphilic block copoly-



mer is determined by the pyrene fluorescence probe method. The graph below shows the pyrene I_1/I_3 excitation ratio plotted against $\log[\text{polymer}]$ (mg/mL).



In this pyrene fluorescence CMC assay, the CMC is identified as:

- (A) The inflection point in the I_1/I_3 versus $\log[\text{polymer}]$ plot, where the ratio transitions from ~ 1.7 – 1.8 (pyrene in polar aqueous phase) to ~ 1.0 – 1.1 (pyrene partitioned into the non-polar micellar core)
- (B) The polymer concentration at which the I_1/I_3 ratio reaches exactly 1.0 for the first time, confirming 100% transfer of all pyrene molecules into micellar cores
- (C) The concentration at which dynamic light scattering (DLS) first detects particles of 10–100 nm, measured simultaneously with the pyrene fluorescence experiment
- (D) The minimum polymer concentration at which the I_1/I_3 ratio equals 1.8, confirming that all pyrene molecules are still in the aqueous phase below the CMC

Q15. Polyamidoamine (PAMAM) dendrimers are synthesised by iterative divergent growth from a central core, with the number of surface groups doubling with each generation. For a PAMAM dendrimer with an ethylenediamine (EDA) core, how many surface amino ($-\text{NH}_2$) groups does a **Generation 3 (G3)** dendrimer possess?

- (A) 16 surface amino groups



- (B) 64 surface amino groups
- (C) 32 surface amino groups
- (D) 8 surface amino groups

Q16. The “totality of evidence” approach in biosimilar development, based on ICH Q5E and regulatory guidance from FDA/EMA, requires comprehensive head-to-head comparison of the biosimilar and the reference product. At which **THREE analytical/regulatory levels** must this comparison be conducted?

- (A) Genomic sequence of the expression cell line, upstream fermentation process parameters, and final drug product accelerated stability profile only
- (B) Drug substance molecular weight by SDS-PAGE, excipient compatibility by DSC, and a single Phase I pharmacokinetic bridging study only
- (C) Process-related impurity profiles (host cell proteins, DNA), container-closure integrity testing, and post-marketing adverse event data only
- (D) Physicochemical/analytical (primary structure, higher-order structure, charge variants, aggregation), biological/functional (receptor binding, potency, Fc effector function), and clinical (PK/PD similarity, safety, immunogenicity, and residual efficacy bridging if needed)

Q17. Under the US Orphan Drug Act (1983) and its implementing regulations (21 CFR Part 316), which criterion **PRIMARILY** qualifies a disease or condition for orphan drug designation?

- (A) The disease must lack any currently approved pharmacological treatment, regardless of patient prevalence; unmet need alone is sufficient for orphan designation
- (B) The disease or condition affects fewer than 200,000 persons in the United States, or there is no reasonable expectation of recovering development costs from US sales alone
- (C) The sponsor must submit Phase II clinical proof-of-concept data before orphan designation can be granted, confirming that the drug is



genuinely promising before development benefits are awarded

- (D) The disease must be a monogenic (single-gene) inherited disorder; acquired diseases such as infections and cancers are ineligible for orphan designation

Q18. Breakthrough Therapy Designation (BTD), introduced by the FDA Safety and Innovation Act (2012), provides enhanced FDA development support. What feature **PRIMARY distinguishes** BTD from Fast Track Designation?

- (A) BTD requires preliminary *clinical* evidence demonstrating substantial improvement over available therapy on at least one clinically significant endpoint, whereas Fast Track requires only that the drug treats a serious condition with potential to address an unmet need (preclinical or early Phase I data may suffice; clinical superiority evidence is not required)
- (B) BTD is available exclusively for biologics and cell/gene therapies, whereas Fast Track Designation is restricted to small-molecule drugs and cannot be awarded to biologics
- (C) BTD guarantees FDA approval within 6 months of NDA/BLA submission, whereas Fast Track provides only a 12-month priority review clock
- (D) BTD replaces the Phase III confirmatory trial requirement; FDA accepts Phase II data alone as sufficient for approval of breakthrough-designated medicines

Q19. A Risk Evaluation and Mitigation Strategy (REMS) for a highly teratogenic oral retinoid requires that prescribers be registered and certified, dispensing pharmacies be enrolled in the programme, and female patients of childbearing potential complete monthly negative pregnancy tests with documented enrolment in a patient registry. Which REMS component specifically encompasses these prescriber certification, pharmacy certification, and patient monitoring requirements?

- (A) Medication Guide (MedGuide) — a written patient-information leaflet



distributed at dispensing that describes the drug's risks including teratogenicity warnings

- (B) Communication Plan (CP) — a targeted letter to healthcare providers explaining the REMS requirements and prescribing conditions at product launch
- (C) Elements to Assure Safe Use (ETASU) — the most restrictive REMS component, which may mandate prescriber or pharmacist certification, patient enrolment in a registry, required laboratory monitoring, and dispensing only through certified facilities
- (D) REMS Assessment Schedule — the periodic FDA submission evaluating whether the programme's goals are being met and whether the strategy remains appropriately calibrated to the risk

Q20. A pharmaceutical company seeks approval for an extended-release (ER) capsule formulation of an already-approved immediate-release (IR) drug. They choose the 505(b)(2) NDA pathway. Compared to a full 505(b)(1) NDA, the **PRIMARY regulatory advantage** of the 505(b)(2) pathway for this ER formulation is:

- (A) The 505(b)(2) pathway waives all bioequivalence and pharmacokinetic bridging studies, relying solely on the IR product's approved labelling to support the ER capsule without any sponsor-conducted clinical work
- (B) The 505(b)(2) pathway allows the ER product to be marketed without any patent certification, automatically bypassing the 30-month Hatch-Waxman stay
- (C) The 505(b)(2) pathway automatically grants 5-year new chemical entity (NCE) exclusivity to the ER formulation even after the IR drug's NCE exclusivity has expired
- (D) The 505(b)(2) pathway permits the applicant to rely on published literature and/or FDA's prior findings of safety and effectiveness for the reference listed drug (RLD), avoiding the need to duplicate the full preclinical and clinical programme already conducted for the approved IR drug



- Q21.** A Phase III randomised controlled trial (RCT) is designed to evaluate a new antihypertensive agent versus placebo. A biostatistician explains that computer-generated block randomisation is essential to the scientific validity of the study. The **PRIMARY statistical purpose** of randomisation in this context is:
- (A) To ensure that patients with more severe hypertension are preferentially allocated to the active treatment arm, so that the therapy is tested in the population most likely to benefit
 - (B) To eliminate selection bias by ensuring each eligible participant has a pre-specified probability of assignment to each treatment group, thereby distributing both known and unknown confounding variables approximately equally between the active and placebo arms
 - (C) To guarantee that double-blinding is maintained throughout the trial, since randomisation codes serve as the primary mechanism for concealing treatment assignment from investigators and participants
 - (D) To satisfy the ICH E9(R1) estimand framework requirement for a minimum enrolment of 500 subjects per arm, which is achievable only through randomisation-based allocation
- Q22.** According to ICH E6(R2) Guideline for Good Clinical Practice (revised 2016), at what point in the clinical trial process must informed consent **FIRST** be obtained from a prospective trial participant?
- (A) Before any trial-specific screening procedures or interventions are performed, including blood tests, imaging, questionnaires, or physical examinations conducted solely to determine eligibility for the trial
 - (B) Consent may be obtained concurrently with the first administration of investigational product, provided the subject has independently reviewed the written consent form beforehand
 - (C) Consent may be deferred until after the run-in period, provided the subject has given verbal assent and the IRB/IEC has approved a deferred consent protocol
 - (D) Within 24 hours of randomisation; ICH E6(R2) permits a brief post-



randomisation window for formal written consent to be completed

- Q23.** According to ICH E2C(R2), which specifies the format and timing of Periodic Benefit-Risk Evaluation Reports (PBRERs), how frequently must a PBRER be submitted during the **THIRD year** after the drug's International Birth Date (IBD)?
- (A) Every 6 months (semi-annually), the same frequency required during the first 2 years after the IBD
 - (B) Every 3 years, as the submission schedule immediately reverts to the long-cycle schedule after year 2
 - (C) Annually (once per year); years 3 and 4 post-IBD require annual PBRER submission under ICH E2C(R2)
 - (D) A PBRER is not required in year 3 unless the FDA issues a written request triggered by a newly identified safety signal
- Q24.** A pharmaceutical company submits confidential manufacturing specifications, composition, and safety data to the US FDA covering the Type I borosilicate glass vials, rubber stoppers, and aluminium crimp caps used in a sterile injectable drug product. According to 21 CFR Part 314.420, this submission is classified as a Drug Master File (DMF) Type:
- (A) Type II — which covers drug substances (APIs) and intermediates used in the preparation of the drug product
 - (B) Type IV — which covers excipients, colorants, flavours, and materials used in the preparation of the drug product
 - (C) Type I — which covers manufacturing site(s), facilities, operating procedures, and personnel
 - (D) Type III — which specifically covers packaging materials, including primary container-closure systems such as vials, stoppers, and closures for injectable drug products
- Q25.** A pharmaceutical company files a New Drug Application (NDA) using the Common Technical Document (CTD) format required by ICH M4. In which CTD Module is the **complete quality (CMC) data** for both the



drug substance and the drug product — including manufacture, characterisation, specifications, stability, and the container-closure system — located?

- (A) Module 3 (Quality), which contains section 3.2.S (drug substance), section 3.2.P (drug product), and section 3.2.A (appendices) with all chemistry, manufacturing, and controls (CMC) data
- (B) Module 2 (CTD Summaries), which contains the Quality Overall Summary (QOS) as a standalone document constituting the complete CMC submission
- (C) Module 5 (Clinical Study Reports), where CMC data are cross-referenced to support the clinical pharmacology sections
- (D) Module 4 (Nonclinical Study Reports), which integrates CMC data with pharmacology, pharmacokinetics, and toxicology data from pre-clinical studies



Detailed Solutions

Q1.

Solution

Concept — Ionizable Lipids in siRNA-LNPs: The defining design criterion for the ionizable lipid in a systemic siRNA-LNP is a pK_a of ~ 6.2 – 6.8 . This range gives the lipid two distinct charge states that are each optimal for one stage of delivery.

Step 1 — Behaviour at physiological pH (blood, pH 7.4): At pH 7.4, which is about 1 pH unit above the $pK_a \approx 6.5$, the Henderson-Hasselbalch relationship predicts that the ionizable amine is predominantly deprotonated (neutral). A neutral lipid minimises electrostatic interaction with negatively charged plasma proteins (opsonins), RBCs, and endothelial surfaces. This prolongs circulation half-life and reduces off-target toxicity and haemolysis compared to a permanently cationic lipid.

Step 2 — Behaviour at endosomal pH (~ 5.5): After receptor-mediated endocytosis, the endosome is acidified to $pH \approx 5.5$ (1 pH unit below the pK_a). At this pH the amine is $> 90\%$ protonated, making the lipid cationic. Cationic lipids form ion pairs with anionic endosomal phospholipids (phosphatidylserine, phosphatidylethanolamine), destabilising the endosomal bilayer and enabling escape of the siRNA cargo into the cytoplasm, where it is loaded onto RISC.

Why other options are wrong:

- Option A: Correct — neutral at pH 7.4, cationic at endosomal pH 5.5, enabling endosomal escape. **Correct.**
- Option B: Permanently cationic lipids (e.g., DOTMA) would cause rapid clearance, haemolysis, and toxicity — precisely the problem ionizable lipids were designed to solve.
- Option C: Maximal protonation at pH 7.4 would make the LNP highly cationic in blood, leading to aggregation with serum proteins and immediate clearance; this is the opposite of the design goal.
- Option D: The ionizable lipid does not undergo irreversible hydrolysis; it is an acid-labile amphiphile whose charge state changes reversibly with pH.

Final Answer: $pK_a \approx 6.5$ gives neutral LNP in blood (pH 7.4) and cationic LNP in endosomes (pH 5.5) for endosomal escape $\Rightarrow$ **A**

Answer: (A) [Go Back to Q1](#)



Q2.

Solution

Concept — Abraxane: Albumin-Mediated Active Tumour Targeting: Abraxane (nab-paclitaxel, ABI-007) exploits endogenous albumin transport pathways to achieve tumour accumulation that exceeds passive EPR alone.

Step 1 — gp60-mediated transcytosis: The glycoprotein gp60 (albondin) is expressed on tumour vascular endothelial cells. Albumin binding to gp60 triggers caveolin-1-mediated endocytosis and vesicular transcytosis of albumin across the endothelial cell, delivering albumin (and bound paclitaxel) into the tumour interstitium. This *active* transcytosis supplements the passive EPR effect.

Step 2 — SPARC-mediated tumour accumulation: SPARC (secreted protein acidic and rich in cysteine, also called osteonectin) is overexpressed in the stroma of many solid tumours (breast, pancreatic, lung). SPARC binds albumin with high affinity, concentrating albumin-bound paclitaxel in the tumour extracellular matrix and providing a drug depot. Tumours with higher SPARC expression show greater response to Abraxane.

Why other options are wrong:

- Option A: The EPR effect alone does not explain the substantially improved tumour accumulation; active transcytosis via gp60/SPARC is the principal mechanism.
- Option B: Active albumin transcytosis via gp60 receptor and SPARC-mediated tumour sequestration. **Correct.**
- Option C: Cremophor EL does not directly mediate P-gp efflux; the mechanism of improvement is not P-gp inhibition.
- Option D: Abraxane does not activate Fc γ receptors; albumin is not an immunoglobulin and does not engage macrophage Fc receptors.

Final Answer: Albumin transcytosis via gp60 $\rightarrow$ SPARC binding in tumour stroma
 $\Rightarrow$ B

Answer: (B) [Go Back to Q2](#)



Q3.

Solution

Concept — ADC Drug-to-Antibody Ratio (DAR) and Pharmacokinetics: The DAR is the average number of drug molecules conjugated per antibody. It critically determines the therapeutic window of an ADC.

Step 1 — Optimal DAR range: Optimal DAR is typically 2–4 for site-specific conjugation and ~ 4 (range 2–8) for stochastic (lysine/cysteine) conjugation. ADCs with $\text{DAR} \leq 2$ have insufficient cytotoxicity; ADCs with $\text{DAR} \geq 8$ suffer from hydrophobic overloading.

Step 2 — Consequences of $\text{DAR} > 8$: Each cytotoxic payload (e.g., MMAE, DM1) is a hydrophobic small molecule. Conjugating > 8 hydrophobic molecules per antibody increases the overall hydrophobicity of the conjugate dramatically: (i) **Aggregation:** hydrophobic interactions drive intermolecular aggregation; (ii) **Rapid MPS clearance:** aggregated and hydrophobic ADCs are recognised and cleared by mononuclear phagocytes, reducing half-life from weeks to hours; (iii) **Increased off-target toxicity:** premature payload release from unstable high-DAR ADCs in non-tumour tissues causes dose-limiting toxicities.

Why other options are wrong:

- Option A: Steric blockage of antigen binding occurs only if payload is conjugated directly in the CDR; this is not the predominant problem with high DAR.
- Option B: High DAR does not enhance antigen-binding affinity; a hook effect is a concentration-dependent immunoassay artifact, not a DAR-related PK event.
- Option C: Rapid clearance due to hydrophobic aggregation and off-target toxicity from premature drug release. **Correct.**
- Option D: DAR does not specifically abolish ADCC; Fc effector function is preserved unless conjugation occurs at Fc glycans (specific site-directed conjugation strategies).

Final Answer: $\text{DAR} > 8 \Rightarrow$ hydrophobic aggregation $\Rightarrow$ rapid clearance and off-target toxicity $\Rightarrow$

Answer: (C) [Go Back to Q3](#)



Q4.

Solution

Concept — CdSe Quantum Dot Toxicity: Cadmium Ion Release: CdSe quantum dots rely on quantum confinement for size-tunable fluorescence. The ZnS shell reduces — but does not fully prevent — core degradation.

Step 1 — Source of Cd^{2+} : Multiple degradation pathways expose the CdSe core: (i) **Photooxidation:** UV irradiation generates reactive oxygen species that oxidise Se^{2-} and dissolve the CdSe lattice, releasing Cd^{2+} through pinholes in the ZnS shell; (ii) **Mechanical stress:** sonication or vigorous mixing can damage the shell; (iii) **Intracellular metabolism:** lysosomal enzymes and the low pH of lysosomes accelerate dissolution of the CdSe core over time.

Step 2 — Cd^{2+} toxicity mechanisms: Free Cd^{2+} displaces zinc from zinc-finger proteins and metallothioneins, inhibits mitochondrial complex I and complex III, uncouples oxidative phosphorylation, and induces excessive reactive oxygen species production. At higher concentrations, mitochondrial membrane potential collapses, triggering intrinsic apoptosis. This is the principal cytotoxicity pathway of CdSe QDs.

Why other options are wrong:

- Option A: ZnS shell does not react with glutathione to form toxic zinc-thiolate complexes under biological conditions; Zn^{2+} released from ZnS is much less toxic than Cd^{2+} .
- Option B: Non-specific protein adsorption (protein corona) is a general nanomaterial phenomenon but does not cause the cytotoxicity characteristic of CdSe QDs.
- Option C: Quantum confinement affects the optical band gap for photon emission; it does not generate ionising radiation.
- Option D: Cd^{2+} release via photooxidation/metabolism is the primary mechanism. **Correct.**

Final Answer: CdSe QD cytotoxicity = Cd^{2+} release $\rightarrow$ ROS generation and mitochondrial dysfunction $\Rightarrow$ D

Answer: (D) [Go Back to Q4](#)



Q5.

Solution

Concept — SPIONs as T2 MRI Contrast Agents: SPIONs (superparamagnetic iron oxide nanoparticles, e.g., Fe_3O_4) are classified as T_2/T_2^* (transverse relaxation) contrast agents producing negative (dark) contrast on T2-weighted MRI images.

Step 1 — Superparamagnetism: Above the magnetic blocking temperature, SPIONs exhibit superparamagnetism: they magnetise strongly in an applied field B_0 but have zero remnant magnetisation in the absence of the field. Within an MRI scanner, each SPION generates a large local dipolar magnetic field extending into the surrounding tissue.

Step 2 — T2 relaxation mechanism: Protons in water molecules diffusing near a magnetised SPION experience rapidly fluctuating local fields (due to diffusion in the inhomogeneous field). These fluctuations cause dephasing of transverse proton magnetisation, accelerating T_2 (spin–spin) and T_2^* relaxation. The result is signal loss on T2-weighted pulse sequences, producing a dark (hypointense) region in tissue where SPIONs accumulate. This is quantified by the r_2 relaxivity ($\text{mM}^{-1}\text{s}^{-1}$).

Why other options are wrong:

- Option A: Magnetic hyperthermia generates heat from Néel/Brownian relaxation under AC fields; this is a therapeutic application, not the MRI contrast mechanism.
- Option B: Accelerated T_2/T_2^* relaxation via local field inhomogeneities $\Rightarrow$ signal loss (dark contrast). **Correct.**
- Option C: SPIONs do not absorb RF pulses; RF pulse absorption is the basis of tissue heating in RF ablation, not MRI contrast.
- Option D: SPIONs are superparamagnetic, not ferrimagnetic; they do not permanently align proton spins or eliminate transverse magnetisation throughout the volume.

Final Answer: SPIONs create local ΔB inhomogeneities $\rightarrow$ accelerated T2 relaxation $\rightarrow$ signal loss = dark contrast on T2-weighted MRI $\Rightarrow$ **B**

Answer: (B) [Go Back to Q5](#)



Q6.

Solution

Concept — PNIPAM Thermoresponsive Polymer: LCST Behaviour: Poly(*N*-isopropylacrylamide) (PNIPAM) has a lower critical solution temperature (LCST) of $\sim 32^\circ\text{C}$ in pure water, tuneable to 34°C by co-monomer selection.

Step 1 — Below LCST ($<34^\circ\text{C}$): At temperatures below the LCST, hydrogen bonding between the carbonyl and amide groups of PNIPAM and surrounding water molecules dominates. The polymer chain adopts an extended, hydrated *coil* conformation. The system is a free-flowing aqueous solution that can be loaded into a syringe and injected.

Step 2 — Above LCST ($>34^\circ\text{C}$): At 37°C (body temperature), which exceeds the LCST, the isopropyl methyl groups become dominant through entropic dehydration; hydrogen bonds with water break, exposing hydrophobic groups that drive rapid chain collapse. The polymer transitions to a compact, dehydrated *globule*, expelling water and forming a physical gel depot at the injection site. This sol-gel transition is reversible upon cooling below the LCST.

Why other options are wrong:

- Option A: Extended hydrophilic coil below LCST $\Rightarrow$ free-flowing injectable solution. **Correct.**
- Option B: Collapse to a globule occurs *above* the LCST, not below it.
- Option C: PNIPAM gelation is a physical (non-covalent) phase transition; irreversible covalent cross-linking is not involved in the LCST mechanism.
- Option D: PNIPAM dissolves readily in water below its LCST; it does not require organic solvents and is not insoluble below 34°C .

Final Answer: Below LCST: PNIPAM is hydrophilic, extended coil, free-flowing injectable solution $\Rightarrow$ A

Answer: (A) [Go Back to Q6](#)



Q7.

Solution

Concept — pH-Responsive Polymers and the Tumour Warburg Effect: Tumour cells preferentially use glycolysis even in the presence of adequate oxygen (the Warburg/aerobic glycolysis effect). The end product, lactic acid (and carbonic acid from CO_2), accumulates in the tumour interstitium, reducing the extracellular pH to 6.4–6.8, compared to ~ 7.4 in normal tissue.

Step 1 — Acidic tumour microenvironment: The Warburg effect drives production of ~ 40 mmol/L lactic acid per hour per litre of solid tumour. This proton excess reduces the tumour extracellular pH (pHe) to 6.4–6.8, creating a ΔpH of 0.6–1.0 units relative to normal tissue (pH 7.4) and blood.

Step 2 — Polymer ionisation and drug release: Poly-methacrylic acid (PMAA), polyacrylic acid, and poly(β -amino ester) chains contain ionisable acidic groups (carboxyl $\text{p}K_a \approx 5\text{--}6$). At blood pH 7.4, these groups are fully ionised and repel each other, but the nanoparticle is still compact due to cross-linking. At tumour pH 6.5–6.8, the additional protonation changes the local electrostatic balance, causing hydrogel swelling and mesh expansion, releasing encapsulated drug preferentially at the tumour site.

Why other options are wrong:

- Option A: Alkaline phosphatase is not an intratumoral pH-raising enzyme; tumour pH is *lower* than normal tissue, not higher.
- Option B: pH-responsive PMAA polymers are stable in the stomach (pH 1.5–3.5); they are not triggered by gastric pH, and the mechanism is not spontaneous hydrolysis.
- Option C: Warburg effect $\rightarrow$ lactic acid $\rightarrow$ tumour pHe 6.4–6.8 $\rightarrow$ polymer ionisation/swelling $\rightarrow$ drug release. **Correct.**
- Option D: Glutathione-responsive release involves disulfide bond cleavage (a separate intracellular stimulus); the question describes extracellular pH response, which is mechanistically distinct.

Final Answer: Warburg effect $\rightarrow$ excess lactic acid $\rightarrow$ extracellular tumour pH 6.4–6.8 $\rightarrow$ polymer ionisation $\rightarrow$ drug release $\Rightarrow$ C

Answer: (C) [Go Back to Q7](#)



Q8.

Solution

Concept — Pulsatile Drug Delivery and Chronopharmacology: Chronopharmacology recognises that disease severity follows circadian (24-hour) rhythms driven by the hypothalamic clock, hormonal cycles, and autonomic tone. Matched drug delivery at the time of peak disease severity maximises efficacy and minimises total dose.

Step 1 — Circadian pattern of nocturnal asthma: Peak bronchoconstriction in nocturnal/early-morning asthma occurs between 3–5 am. Contributing factors include: (i) nadir of circulating cortisol (anti-inflammatory) at ~midnight; (ii) peak airway inflammation and mast cell degranulation in the early morning; (iii) reduced sympathetic tone at night. FEV₁ typically reaches its lowest point at approximately 4 am.

Step 2 — Lag time calculation: Bedtime administration: ~10 pm. Required drug release: ~3–4 am. Lag time needed: ≈ 6 hours. An erodible HEC (hydroxyethyl-cellulose) outer coat erodes over exactly this time, then the inner immediate-release core dissolves rapidly, delivering a salbutamol burst at the time of greatest physiological need.

Why other options are wrong:

- Option A: Salbutamol is not acid-labile; acid protection is not the rationale.
- Option B: Azoreductase is a colonic enzyme used in colon-targeting systems for drugs like mesalazine; pulsatile asthma systems target systemic absorption, not colonic delivery.
- Option C: Salbutamol's half-life is ~3–6 hours, not 30 minutes; even if it were, accumulation avoidance is not the primary design rationale.
- Option D: 6-hour post-bedtime lag = drug burst at ~4 am = peak bronchoconstriction period. Chronopharmacological matching. **Correct.**

Final Answer: $10 \text{ pm} + 6 \text{ h} = 4 \text{ am} = \text{peak nocturnal bronchoconstriction} \Rightarrow \text{circadian synchronisation} \Rightarrow \boxed{\text{D}}$

Answer: (D) [Go Back to Q8](#)



Q9.

Solution

Concept — High-Density Sinking Gastroretentive Systems: High-density gastroretentive systems use heavy excipients to make pellets or tablets denser than gastric contents, causing them to sink to the lowest part of the stomach and resist gastric emptying.

Step 1 — Density comparison: Gastric fluid density $\approx 1.004 \text{ g/cm}^3$. High-density pellets: 2.7 g/cm^3 (achieved with BaSO_4 , density 4.48 g/cm^3 ; ZnO or Fe_2O_3 are also used). These pellets sink immediately to the most dependent region of the stomach.

Step 2 — Gastric retention mechanism: The stomach's most dependent region is the antrum (lower curvature) in the upright position. High-density pellets settle there by sedimentation. The narrow pyloric canal and the sweeping action of migrating motor complexes (MMC) preferentially empty lighter objects; heavy pellets resist this emptying. Unlike floating systems (which require a fed state to elevate gastric fluid volume), high-density systems are retained in both fed and *fasted* states.

Why other options are wrong:

- Option A: BaSO_4 is not paramagnetic (it is diamagnetic); magnetic gastroretention is a separate experimental concept using iron oxide pellets with external magnets.
- Option B: Sedimentation to the antrum/lower curvature due to higher density than gastric fluid. **Correct.**
- Option C: BaSO_4 is insoluble in dilute HCl ; it does not react to release Ba^{2+} or form a gel under gastric conditions.
- Option D: Osmotic-pressure-driven swelling is the mechanism for osmotic gastroretentive systems, not high-density pellets; BaSO_4 is not an osmotic agent.

Final Answer: High-density pellets (2.7 g/cm^3) sink to stomach antrum, retained by gravity and anatomy $\Rightarrow$ **B**

Answer: (B) [Go Back to Q9](#)



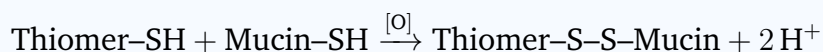
Q10.

Solution

Concept — Thiomers: Covalent Disulfide Mucoadhesion: Conventional mucoadhesive polymers (Carbopol, HPMC, HEC) rely on physical mechanisms: chain entanglement with mucin, hydrogen bonding, and electrostatic interaction. These non-covalent bonds are reversible and weaker than covalent bonds.

Step 1 — Mucin chemistry: Mucin glycoproteins contain cysteine-rich subdomains (D3 subdomain in MUC5B, C-termini in MUC2) where cysteine residues form the inter-chain disulfide cross-links that give mucus its viscoelastic gel structure. Free cysteine residues (–SH) are also present in the mucus layer.

Step 2 — Thiomer–mucin covalent bond: The thiol groups (–SH) of thiomers (e.g., polycarbophil-cysteine) react with free cysteine –SH groups of mucin glycoproteins by oxidative disulfide bond formation (thiol-disulfide exchange) under mild oxidative conditions at physiological pH:



This covalent (–S–S–) bond is ~10-fold stronger than the physical interactions of non-thiolated polymers, providing prolonged mucosal residence times. Additional benefit: thiol groups inhibit P-glycoprotein and MRP2 efflux transporters by depleting ATP (thiol-mediated ATPase inhibition).

Why other options are wrong:

- Option A: Covalent disulfide bonds with mucin cysteine residues. **Correct.**
- Option B: Calcium chelation by thiols is not the primary mucoadhesion mechanism; calcium chelators such as EDTA use carboxylate groups, not thiols.
- Option C: Histidine-thiol reactions (thiol-imidazole) are not a recognised mucoadhesion mechanism in the literature.
- Option D: Tight-junction opening by thiol groups is a paracellular permeation enhancer effect, not the mucoadhesion mechanism; it is a secondary benefit of some thiomers, not the explanation for superior mucoadhesion.

Final Answer: Thiomers form covalent –S–S– bonds with mucin Cys residues ⇒ superior mucoadhesion ⇒ **A**

Answer: (A) [Go Back to Q10](#)



Q11.

Solution

Concept — NLC vs SLN: Crystal Structure and Drug Expulsion: SLNs use a solid lipid (e.g., glyceryl monostearin, stearic acid) as the matrix. During preparation, lipid is melted and solidifies on cooling. The critical problem arises during storage.

Step 1 — SLN limitation: polymorphic transition and drug expulsion: When lipids solidify, they initially form an amorphous or α -crystal structure. Over time (days to weeks), the unstable polymorphic forms convert to the most stable, ordered β -crystal form (lowest free energy). In the perfect β -crystal lattice, drug molecules cannot be accommodated in the rigid crystal structure and are expelled to the nanoparticle surface or into the continuous phase. This causes *drug expulsion*, crystal growth (Ostwald ripening), particle aggregation, and loss of controlled-release properties.

Step 2 — NLC solution: liquid lipid creates imperfect crystal: NLC incorporates a liquid lipid (e.g., oleic acid, isopropyl myristate, Miglyol) that is spatially incompatible with the ordered crystal structure of the solid lipid. The mixture creates an amorphous or nanostructured (multiple crystal imperfections) matrix where drug molecules are accommodated in the defect sites. The imperfect lattice cannot complete the β -transition, preventing drug expulsion and maintaining physical stability over months of storage.

Why other options are wrong:

- Option A: NLC uses a lipid matrix, not PLGA polymer; PLGA is a separate (polymeric nanoparticle) delivery system with different degradation mechanisms.
- Option B: A second surfactant layer is not the defining feature of NLC; both SLN and NLC use similar surfactant stabilisation.
- Option C: Imperfect crystal structure from solid+liquid lipid blend prevents drug expulsion. **Correct.**
- Option D: Preparation method does not define NLC vs SLN; both can be prepared by hot HPH; NLC is defined by its structural composition.

Final Answer: Solid + liquid lipid blend $\rightarrow$ imperfect crystal $\rightarrow$ no drug expulsion
 $\Rightarrow$

Answer: (C) [Go Back to Q11](#)



Q12.

Solution

Concept — Transferosomes: Transepidermal Water Gradient as Driving Force: Transferosomes (elastic vesicles, Cevc & Blume 1992) penetrate intact skin through intercellular lipid channels of the stratum corneum. Their penetration requires a driving force; this is provided by the transepidermal osmotic/water gradient.

Step 1 — The transepidermal water gradient: The viable epidermis and dermis have a water activity of $\sim 100\%$ (fully hydrated). At the skin surface, continuous transepidermal water loss (TEWL) evaporates water, creating a water activity/concentration gradient from inside ($\sim 100\%$) to the surface ($< 100\%$). This gradient drives net movement of water outward and, by osmotic coupling, drives any deformable vesicle applied to the surface to migrate *inward* (down the osmotic gradient toward higher water activity).

Step 2 — Consequence of occlusion: When an impermeable patch or dressing is placed over the skin, evaporation of water from the skin surface is blocked. The skin surface becomes as hydrated as the underlying dermis, eliminating the water concentration gradient. Without the gradient, there is no osmotic driving force for inward vesicle migration. Transferosomes do not penetrate under occlusion, unlike oil-based vehicles that depend on concentration gradients for passive diffusion.

Why other options are wrong:

- Option A: Transferosomes are typically formulated in aqueous buffers without high residual organic solvent; solvent burn is not the reason for non-occlusion.
- Option B: The edge activator mechanism is about bilayer elasticity enabling squeezing, not about desorption under occlusion.
- Option C: Fusion with sebaceous gland secretions is not a described mechanism; sebaceous glands occupy $< 1\%$ of skin surface area.
- Option D: Occlusion eliminates the transepidermal water gradient $\Rightarrow$ no driving force for vesicle permeation. **Correct.**

Final Answer: Non-occlusion $\rightarrow$ transepidermal water gradient maintained $\rightarrow$ osmotic driving force for vesicle penetration $\Rightarrow$ **D**

Answer: (D) [Go Back to Q12](#)



Q13.

Solution

Concept — Exosomes as Drug Carriers: Biological Advantages: Exosomes are 30–150 nm extracellular vesicles secreted by virtually all cell types when multi-vesicular endosomes fuse with the plasma membrane. Their lipid bilayer and surface proteome reflect the parent cell.

Step 1 — Natural surface proteins: Exosome surfaces display: (i) tetraspanins (CD9, CD63, CD81) involved in intercellular communication; (ii) integrins that guide tissue homing (e.g., $\alpha_v\beta_3$ directs exosomes to bone, $\alpha_6\beta_4$ to lung); (iii) CD47 (“integrin-associated protein”), a “don’t-eat-me” signal that inhibits phagocytosis by macrophages by engaging SIRP α on macrophages, analogous to PEGylation of synthetic vesicles.

Step 2 — Immunological compatibility: Because exosomes are composed of endogenous membrane components, autologous exosomes (from the patient’s own cells) are not recognised as foreign by the immune system. Even allogeneic exosomes carry lower immunogenicity than synthetic liposomes due to the presence of natural self-recognition proteins. This reduces complement activation, opsonisation, and rapid clearance.

Why other options are wrong:

- Option A: Exosomes are notoriously difficult to scale up; cell culture yields are low (micrograms per litre), and drug loading efficiency is generally worse than liposomes.
- Option B: Natural cellular homing and intrinsically low immunogenicity. **Correct.**
- Option C: Exosome membranes consist of biological phospholipids, sphingolipids, and cholesterol from the parent cell — not synthetic PEGylated lipids.
- Option D: Exosome luminal pH is not maintained at 5.0; that is the pH of late endosomes/lysosomes from which they are derived, but secreted exosomes equilibrate with extracellular pH (~ 7.4).

Final Answer: Exosomes have endogenous homing proteins (integrins, CD47) and natural self-recognition $\rightarrow$ lower immunogenicity $\Rightarrow$ **B**

Answer: (B) [Go Back to Q13](#)



Q14.

Solution

Concept — CMC Determination by Pyrene Fluorescence: Pyrene is an environmentally sensitive fluorophore with distinct excitation band peaks: $I_1 \approx 373$ nm (vibronic band 1, polarity-sensitive) and $I_3 \approx 384$ nm (vibronic band 3, polarity-insensitive). The I_1/I_3 ratio reports on local polarity.

Step 1 — Pyrene below CMC: Below the CMC, the polymer exists as individual unimer chains in aqueous solution. Pyrene remains in the polar aqueous phase, experiencing a hydrophilic environment. The I_1/I_3 ratio is ≈ 1.7 – 1.8 , characteristic of a polar solvent.

Step 2 — Pyrene above CMC: Above the CMC, micelles form with a hydrophobic core. Pyrene is highly lipophilic ($\log P \approx 5$) and partitions preferentially into the non-polar micellar core. In this hydrophobic environment, the I_1/I_3 ratio drops to ≈ 1.0 – 1.1 , characteristic of non-polar solvents (e.g., heptane). The CMC is identified as the inflection point in the I_1/I_3 versus $\log[\text{polymer concentration}]$ sigmoidal curve — the midpoint of the ratio transition from high to low values.

Why other options are wrong:

- Option A: Inflection point (midpoint of sigmoid) in I_1/I_3 vs $\log[C]$ curve. **Correct.**
- Option B: The ratio asymptotically approaches 1.0 but does not reach exactly 1.0 (micellar core is not perfectly non-polar, and not all pyrene partitions into micelles); the inflection is the analytical marker, not a strict ratio of 1.0.
- Option C: DLS is an independent technique; the pyrene method alone identifies the CMC by the spectroscopic transition, without DLS.
- Option D: Option D describes the pre-CMC plateau ($I_1/I_3 \approx 1.8$), which extends from very low concentration to the CMC; the CMC is where this plateau *ends* (inflection), not where it begins.

Final Answer: CMC = inflection point in I_1/I_3 vs $\log[\text{polymer}]$ curve (transition from ~ 1.8 to ~ 1.0) $\Rightarrow$ **A**

Answer: (A) [Go Back to Q14](#)



Q15.

Solution

Concept — PAMAM Dendrimer Generations and Terminal Group Number: PAMAM dendrimers grow by divergent synthesis from a central core. Each generation (branch shell) doubles the number of terminal functional groups. With an EDA (ethylenediamine) core providing 4 initiator sites (NH_2 groups), the formula for terminal groups at generation n is:

$$N_{\text{terminal}} = 4 \times 2^n$$

Step 1 — Apply the formula for G3:

$$N_{\text{G3}} = 4 \times 2^3 = 4 \times 8 = 32 \text{ surface amino groups}$$

Step 2 — Verify by generation sequence: G0: $4 \times 2^0 = 4$; G1: $4 \times 2^1 = 8$; G2: $4 \times 2^2 = 16$; G3: $4 \times 2^3 = 32$; G4: $4 \times 2^4 = 64$; G5: $4 \times 2^5 = 128$. (G4 = 64 was covered in Paper 3.)

Why other options are wrong:

- Option A: 16 corresponds to G2 ($4 \times 2^2 = 16$), not G3.
- Option B: 64 corresponds to G4 ($4 \times 2^4 = 64$), not G3.
- Option C: $32 = 4 \times 2^3$. **Correct.**
- Option D: 8 corresponds to G1 ($4 \times 2^1 = 8$), not G3.

Final Answer: G3 PAMAM (EDA core): $N = 4 \times 2^3 = 32$ surface amino groups $\Rightarrow$

C

Answer: (C) [Go Back to Q15](#)

Q16.

Solution

Concept — Biosimilar Totality of Evidence Approach: The totality-of-evidence (TOE) approach, endorsed by the FDA Biosimilar Guidance (2015, revised 2019), EMA CHMP/437/04, and underpinned by ICH Q5E (comparability), requires a stepwise, hierarchical comparison of the biosimilar and reference product.

Step 1 — Level 1: Physicochemical and analytical characterisation: Primary structure (amino acid sequence by peptide mapping, disulfide bonding), higher-order structure (secondary/tertiary structure by CD spectroscopy, HDX-MS, in-



trinsic fluorescence), post-translational modifications (glycan profiling by LC-MS), charge variants (IEF, cation exchange HPLC), aggregation (SEC, DLS, AUC), and purity (RP-HPLC, SDS-PAGE).

Step 2 — Level 2: Biological/functional characterisation: Target receptor binding (SPR, ELISA), Fc effector functions (FcRn binding, ADCC, CDC for antibody biosimilars), cell-based potency assays, and PK/PD similarity in animal models.

Step 3 — Level 3: Clinical characterisation: Comparative PK study (single-dose cross-over in healthy volunteers or patients), safety (immunogenicity, AEs), and, if residual uncertainty remains after analytical and functional data, comparative efficacy study. The TOE reduces the clinical development burden when analytical similarity is robustly demonstrated.

Why other options are wrong:

- Option A: Genomic sequencing and fermentation parameters are development tools, not the three analytical levels of the comparability exercise.
- Option B: SDS-PAGE MW and DSC alone are insufficient for biosimilar characterisation; the comprehensive analytical package is required.
- Option C: Impurity profiles and container-closure testing alone do not constitute the three levels of biosimilar comparison.
- Option D: Physicochemical/analytical, biological/functional, and clinical. **Correct.**

Final Answer: Totality of evidence = analytical + biological/functional + clinical comparison $\Rightarrow$

Answer: (D) [Go Back to Q16](#)

Q17.

Solution

Concept — US Orphan Drug Act (1983) Designation Criteria: The Orphan Drug Act (ODA) was enacted to incentivise development of drugs for rare diseases by providing regulatory and economic incentives.

Step 1 — Prevalence criterion (primary): A disease qualifies for orphan designation if it affects **fewer than 200,000 persons** in the United States at the time of designation (21 CFR §316.20(b)(1)). This is equivalent to a prevalence of ~ 1 in 1,500 Americans.

Step 2 — Cost-recovery criterion (secondary): For diseases with worldwide



prevalence exceeding 200,000 US persons but where sales are expected to be limited (e.g., rare cancers), designation may also be granted if there is **no reasonable expectation of recovering development and production costs from US sales alone** within 7 years of approval.

ODA benefits: 7-year market exclusivity (no FDA approval of same drug for same indication during exclusivity period), 50% tax credit for qualified clinical trial costs, FDA fee waivers, eligibility for fast-track and priority review.

Why other options are wrong:

- Option A: Unmet need alone (without the prevalence criterion) is not sufficient; the prevalence threshold is the fundamental legal criterion.
- Option B: Prevalence <200,000 US persons OR no reasonable cost-recovery expectation. **Correct.**
- Option C: Phase II data are not required for orphan designation; designation is a pre-approval incentive based on disease prevalence, granted as early as pre-IND stage.
- Option D: Many orphan drugs target acquired diseases: cancers (e.g., imatinib for CML at time of designation), rare infections, and autoimmune diseases. The ODA does not restrict to genetic diseases.

Final Answer: Orphan designation = disease affects <200,000 US persons OR no reasonable cost recovery $\Rightarrow$ **B**

Answer: (B) [Go Back to Q17](#)

Q18.

Solution

Concept — FDA Breakthrough Therapy Designation vs Fast Track Designation: Both programmes are expedited development pathways for serious or life-threatening conditions, but they differ significantly in their evidentiary thresholds and scope of FDA support.

Step 1 — Fast Track Designation criteria: Granted if: (1) the drug treats a serious or life-threatening condition AND (2) the drug has the potential to address an unmet medical need. Clinical evidence is *not* required; preclinical data or early Phase I safety data may be sufficient. Benefits: rolling review, more frequent FDA meetings, eligibility for Accelerated Approval and Priority Review.

Step 2 — Breakthrough Therapy Designation criteria: Granted if: (1) serious or life-threatening condition AND (2) **preliminary clinical evidence** indicates that



the drug may demonstrate substantial improvement over available therapy on a **clinically significant endpoint**. The clinical superiority requirement distinguishes BTD from Fast Track. BTD provides all Fast Track benefits plus: intensive guidance from senior FDA cross-disciplinary review team, organisational commitment from FDA senior management, and more frequent meetings at critical development milestones.

Why other options are wrong:

- Option A: BTD requires preliminary clinical evidence of substantial improvement; Fast Track requires only potential to address unmet need. **Correct.**
- Option B: Both BTD and Fast Track are available for any drug (small molecules, biologics, cell/gene therapies); neither is product-class restricted.
- Option C: BTD does not guarantee approval timelines; it provides intensive FDA collaboration, not a fixed review clock.
- Option D: Phase III confirmatory trials remain required for BTD; Phase II data alone may allow Accelerated Approval (using surrogate endpoint) under a separate pathway, but full approval typically requires Phase III.

Final Answer: BTD requires preliminary clinical evidence of superiority; Fast Track requires only potential to address unmet need $\Rightarrow$ A

Answer: (A) [Go Back to Q18](#)

Q19.

Solution

Concept — REMS Components: Elements to Assure Safe Use (ETASU): A Risk Evaluation and Mitigation Strategy (REMS) is mandated by FDA (21 USC §505-1, FDAAA 2007) when the Agency determines that additional risk management beyond standard labelling is necessary to ensure benefits outweigh risks.

Step 1 — REMS components hierarchy:

- Medication Guide (MedGuide):** Written patient information distributed at dispensing. Required in *all* REMS.
- Communication Plan (CP):** Targeted letters or educational materials to healthcare providers. Optional.
- Elements to Assure Safe Use (ETASU):** The most restrictive component. Possible elements include: certified prescriber programme, certified pharmacy network, patient enrolment in registry, required laboratory tests (e.g., ANC for clozapine, pregnancy test for isotretinoin/thalidomide), dispensing



only in certain healthcare settings (e.g., hospital for IV formulations).

(iv) **Implementation System:** Mechanism to ensure ETASU are followed.

(v) **REMS Assessment:** Periodic evaluation submitted to FDA.

Step 2 — Apply to the scenario: Certified prescribers, certified pharmacies, patient enrolment, pregnancy testing with documented enrolment = all ETASU elements. Examples: iPledge (isotretinoin), REMS for thalidomide (THALOMID REMS), RevAssist (lenalidomide), CLOZAPINE REMS (ANC monitoring).

Why other options are wrong:

- Option A: MedGuide is a patient leaflet; it does not restrict who can prescribe or dispense.
- Option B: Communication Plan is an educational outreach tool, not a prescriber or pharmacy restriction mechanism.
- Option C: ETASU encompasses prescriber/pharmacy certification and patient monitoring requirements. **Correct.**
- Option D: REMS Assessment Schedule is the evaluation reporting mechanism, not the component that implements distribution restrictions.

Final Answer: Prescriber/pharmacy certification + patient registry/monitoring = Elements to Assure Safe Use (ETASU) ⇒ C

Answer: (C) [Go Back to Q19](#)

Q20.

Solution

Concept — 505(b)(2) NDA Pathway: Reliance on Prior Data: Section 505(b)(2) of the FD&C Act (21 CFR §314.54) provides a regulatory pathway for drug applications that rely, in whole or in part, on data or information not generated by the applicant to support the safety or effectiveness of the proposed drug.

Step 1 — Nature of the 505(b)(2) application: Unlike a 505(b)(1) NDA (where the applicant conducts and owns all studies) and a 505(j) ANDA (generic, requires only bioequivalence), 505(b)(2) allows the sponsor to cite: (a) published literature; (b) FDA's prior safety/efficacy findings for the reference listed drug (RLD). This means the sponsor does not need to repeat the original pharmacology, toxicology, and clinical studies already conducted for the approved IR formulation, as long as the bridge to the new ER formulation is established through formulation-specific studies (BE/PK bridging, dissolution comparison).



Step 2 — Application to the ER formulation scenario: The sponsor of the ER capsule needs to demonstrate: (i) the ER formulation is bioequivalent to or pharmacokinetically distinct from the IR product in a defined, justified way; (ii) the safety and efficacy of the IR drug (from the existing NDA) support the ER formulation's benefit-risk profile. The sponsor avoids repeating rodent toxicology, human Phase I/II pharmacology studies, and pivotal Phase III trials, saving years and hundreds of millions in development costs.

Why other options are wrong:

- Option A: 505(b)(2) does not waive PK/BE studies; it waives only the studies already conducted for the RLD. Bridging PK studies are still required.
- Option B: Patent certification (Paragraph III or IV) is still required under Hatch-Waxman; 505(b)(2) does not bypass patent challenges.
- Option C: 3-year exclusivity (for a new condition of use, new dosage form) may be available, but not 5-year NCE exclusivity; and it is not automatic.
- Option D: Reliance on published literature and RLD data avoids duplicating the original preclinical/clinical development programme. **Correct.**

Final Answer: 505(b)(2) allows reliance on RLD data/published literature, avoiding full preclinical/clinical development duplication ⇒ D

Answer: (D) [Go Back to Q20](#)

Q21.

Solution

Concept — Randomisation in Phase III RCTs: Elimination of Selection Bias: Randomisation is the cornerstone of the RCT design and the primary mechanism by which causal inference (not merely association) can be drawn from clinical trial data.

Step 1 — What is selection bias? Selection bias occurs when the assignment of participants to treatment groups is influenced (consciously or unconsciously) by the investigator or by participants' preferences, resulting in groups that differ systematically in baseline characteristics. For example, without randomisation, sicker patients might be enrolled preferentially in the treatment arm, confounding the treatment effect estimate.

Step 2 — How randomisation addresses this: By assigning each eligible participant according to a pre-specified probability (e.g., 1:1 for a two-arm trial), randomisation ensures that *both measured confounders* (age, sex, disease severity,



comorbidities) and *unmeasured/unknown confounders* (genetic polymorphisms, lifestyle factors, microbiome) are distributed approximately equally between treatment and placebo groups. This distributional balance is guaranteed *on average* for large samples (by the law of large numbers) and is the statistical basis for valid hypothesis testing in RCTs (ICH E9, ICH E9(R1)).

Why other options are wrong:

- Option A: Preferential allocation of sicker patients to the treatment arm introduces confounding; that is the *opposite* of what randomisation achieves.
- Option B: Elimination of selection bias by equal probability allocation, distributing known and unknown confounders. **Correct.**
- Option C: Blinding (concealing treatment assignment) is maintained by masking (identical placebo) and sealed randomisation codes, but the *purpose* of randomisation is confounder distribution, not blinding itself.
- Option D: ICH E9(R1) specifies estimand principles, not a minimum sample size of 500 per arm; sample size is determined by power calculations based on effect size and variability.

Final Answer: Randomisation eliminates selection bias by distributing known and unknown confounders equally $\Rightarrow$ **B**

Answer: (B) [Go Back to Q21](#)

Q22.

Solution

Concept — ICH E6(R2) GCP: Timing of Informed Consent: Informed consent is a cornerstone of GCP and research ethics, requiring that each participant voluntarily agrees to participate after being fully informed of the nature, risks, benefits, and alternatives of the trial.

Step 1 — ICH E6(R2) requirements (Section 4.8): Section 4.8.1 states: “The subject’s consent must be documented by means of a written, signed and dated informed consent form.” More critically for timing, Section 4.8.2 states that the signed consent must be obtained before the subject participates in the trial. The FDA’s 21 CFR §50.25 mirrors this requirement.

Step 2 — What constitutes “participation”? Any procedure performed solely to determine eligibility for the trial — including blood sampling, physical examination, questionnaires, imaging — constitutes a trial-related activity. Consent must be obtained *before* any such procedure, not merely before randomisation or



dosing. This protects participant autonomy and prevents coercion by allowing potential participants to decline screening.

Why other options are wrong:

- Option A: Before any trial-specific screening procedures, including eligibility assessments. **Correct.**
- Option B: Concurrently with the first dose would be far too late; consent must precede any trial-specific activity, including screening.
- Option C: Deferred consent protocols are limited to emergency situations (e.g., acute MI, trauma) with specific IRB approval; they are not a general provision of ICH E6(R2) for normal phase III trials.
- Option D: A 24-hour post-randomisation window is not recognised in ICH E6(R2); this would make consent retrospective and violate the principle of voluntariness.

Final Answer: Consent required before any trial-specific screening procedures (ICH E6(R2) Section 4.8) ⇒ A

Answer: (A) [Go Back to Q22](#)

Q23.

Solution

Concept — PBRER Submission Schedule under ICH E2C(R2): The Periodic Benefit-Risk Evaluation Report (PBRER, formerly PSUR) is a cumulative safety and benefit-risk evaluation required for marketed products. ICH E2C(R2) (adopted 2012) specifies the data lock point, format, and submission frequency.

Step 1 — ICH E2C(R2) submission schedule from International Birth Date (IBD):

- Years 1–2 post-IBD: **every 6 months** (semiannual; 2 PBRERs per year)
- Years 3–4 post-IBD: **annually** (1 PBRER per year)
- After Year 5 (if benefit-risk profile is stable and agreed with the regulatory authority): **every 3 years**

Step 2 — Year 3 frequency: Year 3 is in the second period (years 3–4), which requires annual PBRER submission. Only one report is due during the third year after the IBD.

Why other options are wrong:



- Option A: 6-monthly frequency applies to years 1–2, not year 3.
- Option B: 3-yearly frequency applies after year 5 (or later, if agreed); it does not begin at year 3.
- Option C: Annually in years 3–4. **Correct.**
- Option D: PBRER submission is obligatory at all stages; FDA does not need to request it unless there is a deviation from the standard schedule.

Final Answer: Year 3 post-IBD = annual PBRER submission (ICH E2C(R2)) ⇒ C

Answer: (C) [Go Back to Q23](#)

Q24.

Solution

Concept — FDA Drug Master Files (DMF) Classification: DMFs are voluntary submissions to FDA that allow confidential disclosure of manufacturing information (facility, process, materials, packaging) to support NDAs, ANDAs, and BLAs without direct disclosure to the applicant who references the DMF.

Step 1 — DMF type definitions (21 CFR §314.420):

- **Type I:** Manufacturing site(s), facilities, operating procedures, personnel
- **Type II:** Drug substance (API), drug substance intermediate, material used in preparation of drug substance
- **Type III:** Packaging material (primary and secondary container-closure systems)
- **Type IV:** Excipient, colorant, flavour, essence, or material used in the preparation of the drug product
- **Type V:** FDA-accepted reference information

Step 2 — Classify the submission: The vials (primary container), rubber stoppers (primary closure), and aluminium crimp caps (primary seal) are all components of the *container-closure system* of the injectable product. These are *packaging materials* ⇒ Type III DMF.

Why other options are wrong:

- Option A: Type II covers the API (drug substance), not the container.
- Option B: Type IV covers excipients and materials used in the drug product formulation, not the final container-closure system.
- Option C: Type I covers manufacturing facilities and processes, not packaging.



- Option D: Type III covers packaging/container-closure materials for drug products. **Correct.**

Final Answer: Vials + stoppers + crimp caps = container-closure system = Type III DMF $\Rightarrow$ D

Answer: (D) [Go Back to Q24](#)

Q25.

Solution

Concept — CTD Format: Module 3 (Quality/CMC): The Common Technical Document (CTD, ICH M4) is the internationally harmonised format for NDA/BLA submissions accepted by FDA, EMA, PMDA, and other ICH member authorities.

Step 1 — CTD Module structure:

- **Module 1:** Administrative information (region-specific: prescribing information, application forms, cover letters)
- **Module 2:** CTD Summaries (2.3 Quality Overall Summary [QOS]; 2.4 Nonclinical Overview; 2.5 Clinical Overview; 2.6 Nonclinical Written Summaries; 2.7 Clinical Summary)
- **Module 3:** Quality — the complete CMC dossier:
 - 3.2.S Drug Substance (3.2.S.1 General Information, 3.2.S.2 Manufacture, 3.2.S.3 Characterisation, 3.2.S.4 Control of Drug Substance, 3.2.S.5 Reference Standards, 3.2.S.6 Container Closure System, 3.2.S.7 Stability)
 - 3.2.P Drug Product (manufacture, excipients, development, closure, specifications, stability)
 - 3.2.A Appendices (facilities, adventitious agents, novel excipients)
- **Module 4:** Nonclinical Study Reports (pharmacology, PK/TK, toxicology)
- **Module 5:** Clinical Study Reports (all clinical trial reports, CSR)

Step 2 — Identify the correct module: All chemistry, manufacturing, and controls (CMC) data — characterisation, manufacture, specifications, stability, and container-closure — for both drug substance and drug product reside in **Module 3**.

Why other options are wrong:

- Option A: Module 3 contains the complete CMC data. **Correct.**



- Option B: Module 2.3 (QOS) is a summary of Module 3; it is not the full CMC dossier.
- Option C: Module 5 contains clinical pharmacology (PK/PD) study reports, not CMC data.
- Option D: Module 4 contains nonclinical (preclinical) pharmacology, PK, and toxicology reports; CMC is in Module 3.

Final Answer: Complete CMC data for drug substance and drug product = CTD Module 3 (3.2.S, 3.2.P, 3.2.A) ⇒

Answer: (A) [Go Back to Q25](#)



Answer Key

Q	Ans	Q	Ans	Q	Ans	Q	Ans	Q	Ans
1	A	2	B	3	C	4	D	5	B
6	A	7	C	8	D	9	B	10	A
11	C	12	D	13	B	14	A	15	C
16	D	17	B	18	A	19	C	20	D
21	B	22	A	23	C	24	D	25	A

